

Short Review on Spectral Methods for the Determination of Chromiun in Human Body Fluids and Tissues

Maria G. Angelova, Atanaska N. Bozhinova

Department of Chemistry and Biochemistry & Physics and Biophysics, University of Medicine-Pleven, Pleven, Bulgaria

Email address

angelovamg@abv.bg (M. G. Angelova)

To cite this article

Maria G. Angelova, Atanaska N. Bozhinova. Short Review on Spectral Methods for the Determination of Chromiun in Human Body Fluids and Tissues. *Open Science Journal of Analytical Chemistry*. Vol. 2, No. 3, 2015, pp. 13-19.

Abstract

Background: The data for the determination of chromium concentrations in biological samples are still contradictory and for some biological objects varied within orders of magnitude. They are not fully clarified the role and participation in the metabolism in healthy subjects and in various diseases and conditions. Aim: One reason for the outstanding problems in the study of the functions of the chromium in the human organism is a need for accurate, with low detection limits, and access to a biomedical laboratory methods for the analytical determination. That's why we set the aim to make a brief review of spectral methods for determination of chromium in human body fluids and tissues. Methods, results: Analytical methods which most commonly to apply at determining the chromium in biomaterials are the atomic spectral (AS) - Flame atomic absorption spectroscopy, Electrothermal atomic absorption spectrometry, Inductively coupled plasma optical emission spectrometry and Inductively coupled plasma mass spectrometry. They are accurate and have lower limits of detection, but not all biomedical laboratories have them. Of the molecular spectral, applicable methods with spectrophotometric indication for determination of chromium in biomaterials are the methods which used suitable chelating agents of chromium in micellar medium. They have limits of detection, comparable to AS-methods and are available to all biomedical laboratories. Conclusions: The lag in specifying the chromium concentrations in human bio-samples, related with clarifying the role of chromium in health and disease, can be overcome using both atomic spectral and spectrophotometric methods. Any study relating to the determination of chromium in human body fluids and tissues will contribute to the analytic of chromium and related the determining his biomedical research.

Keywords

Cr AS-methods Body Fluids, Cr AS-methods Tissues, Cr Photometric Body Fluids, Cr Photometric Tissues

1. Introduction

As one of the essential trace elements-biotics, chromium involved in the biochemical processes in the body. Its content in human body fluids and tissues varies from ng (ng / ml) in the blood serum to μ g range (μ g/g) in the hair [1]. Changes in chromium concentrations were observed in various diseases and conditions, but the involvement of chromium in the exchange is not yet fully elucidated [2, 3]

One of the reasons for delays in studying the function of chromium in the body is the need for accurate, accessible, with low detection limits of the analytical methods for its determination in biological samples. The low chromium concentrations - about three orders of magnitude lower than those of iron, copper and zinc, are a major cause of the more complicated and often inaccurate analytical determination.

In determining the concentration in nanomoles range, chemical methods - classical photometry and electrochemical methods require application of slow, laborious, multi-step methods for pre-concentration and separation, involving the use of large volume and mass of biological samples. The large volume or mass required for these methods, for example 10 or 20 ml of blood serum, making them unsuitable for analysis of most of the biological samples.

Instrumental methods of analysis - AAS (Atomic absorption spectrometry), NAA (Neutron activation analysis), Inductively Coupled plasma Optical Emission Spectrometry (ICP-OES) µ Inductively Coupled plasma mass spectrometry (ICP-MS) have low limits of detection, but require expensive and complex equipment. As with the chemical methods, and at the instrumental, the complicate and variable biological matrix requires the use of prior processing the samples in which the methods are becoming more labor intensive [1 - 6, 7, 8].

Methods for determining the appropriate metal chelating agents in micellar medium can compete with instrumental methods. They have low limits of detection, while photometric indication; these limits are combined with access for scientific and routine technique of execution. [9].

With regard to solving the analytical problems in the determination of chromium in biological samples, we set the aim to make a brief overview of spectral methods for determination of chromium in biological samples. Contemporary values for the distribution of the content of chromium in human body fluids and tissues are given in Table 1.

Samples		Selected contemporary values in healthy people			
Measuring units		*Cavy. ± SD	C _{Cr(from - to)} or C _{av.v}	Reference	
Blood, ET-AAS	μg/l	0.22 ± 0.07	0.08-0.51	[1]	
Blood, ET-AAS	μg/l		0.22-12.00	[4]	
Blood, ET-AAS	μg/l	35.04 ± 26.02		[5]	
Blood, ETAAS	μg/l	59.2 ± 2.94	51.92-63.50	[2]	
Blood, ET-AAS	ug/l	6.19 ± 0.03		[6]	
Blood, ETAAS	ug/l		0.5 - 5.1	[3]	
Blood ISP-MS	μg/l	0.44 ± 0.27	0.01-1.2	[10]	
Blood, ISP-OES	ug/l	0.23 ± 0.03	0.00-0.28	[11]	
Serum, ISP-OES	ug/l	$1,80\pm1,40$		[7]	
Serum, ISP-MS	μg/l		1.0	[8]	
Serum, ET-AAS	μg/l	0.16±0.06	0.08-0.36	[12]	
Serum, ET-AAS	μg/l	0.17 ± 0.05	0.06 - 0.43	[1]	
Serum, ET-AAS	ug/l		12 - 60	[13]	
Serum FAAS	μg/l	0.88 ± 0.2		[14]	
Urine, ET-AAS	ug/l		0.30-0.90	[15]	
Urine, ET-AAS	μg/l		0.24-1.8	[16]	
Urine, FAAS	ug/l		6.10	[17]	
Urine, ISP-OES	ug/l	9.00±7.00		[7]	
Urine, FIA FAAS	ug/l	3.80 ± 0.20	0.04-50	[18]	
Urine, ET-AAS	ug/l	9.20 ± 2.54	7.50-11.40	[2]	
Urine, ET-AAS	ug/l	1.18 ± 0.23		[19]	
Urine, ET-AAS	ug/l	0.32±0.20	0.08-0.90	[12]	
Lung, ET-AAS	ug/g	0.117 ± 0.053	0.00161 - 0.40	[1]	
Milk. ET-AAS	ug/l		16.0	[1]	
Milk, ISP-OES	ug/kg		72	[20]	
Human milk. ET-AAS	ug/l		from 0.69 ± 0.01 to 17.33 ± 0.59	[21]	
Saliva, ET-AAS	μg/l		0.80 - 3.60	[1]	
Skin. ETAAS	ng/g		21.0	[1]	
Sweat, ETAAS	ug/l	2.0 ± 1.2		[1]	
Nails, ISP-OES	ug/ml	2.37±0.64		[7]	
Nails, ISP-OES	ug/g	1.47 ± 0.56		[22]	
Nails. ET-AAS	ug/g		0.8–1.4	[15]	
Nails, ET-AAS	μg/g	1.16±1.05		[16]	
Ervthrocytes, ET-AAS	ug/l		1.4-2.5	[15]	
Ervthrocytes, ET-AAS	ug/l		0.14-4.58, 1.54	[16]	
Ervthrocvtes, ET-AAS	ug/l	3.21 ± 2.20	,	[23]	
Plasma, FAAS	μg/dl	64.98±5.17		[24]	
Liver, blood, FAAS	μg/ml		0.25-6.0	[25]	
Liver, ET-AAS	ppm	2.49±0.89		[26]	
Hair, FAAS	ug/g		0.20	[27]	
Hair, FAAS	mg/g		1.0152 ± 0.8844	[28]	
Hair. FAAS	mg/g	3.97±3.44		[29]	
Hair, ETAAS	μg/g	3.6 ± 0.45	3.05 - 4.2	[2]	
Hair. ISP-OES	ug/ml	4.36±1.03		[7]	
Hair. ISP-OES	ug/g	3.32 ± 0.22		[22]	
Hair. ISP-OES	ug/g	3.32 ± 0.22		[22]	
Hair, ISP-OES	μg/g	0.92 ± 0.65		[11]	
Hair, ET-AAS	μg/g	2.45 ± 1.3		[30]	
Hair, ET-AAS	ng/g	4280 ± 278		[31]	
Hair, ET-AAS	μg/g	1.1±0.3		[16]	
Prostate, NAA-LLR	mg/kg	0.49 ± 0.07		[32]	
Prostate, ISP-MS	mg/kg	0.53 ± 0.08		[32]	
Tissues, ETAAS	μg/l		37.04	[5]	

Table 1. Chromium concentrations (C_{Cr}) in human body fluids and tissues.

Samples		Selected contemporary va		
Measuring units		$C_{av.v.} \pm SD$	C _{Cr(from - to)} or C _{av,v}	Reference
Tissues, ET-AAS	µg∕g	2.84±5.17		[5]
Bones, FAAS	µg/g	7.87±11.92		[33]
Urine, ET-AAS	μg/g		0.08	[34]
Hip joint tissues, ETAAS	μg/g		5.33 - 17.86	[35]

 $C_{avv} \pm SD$ - Cr concentration average value \pm standard deviation

Table 1 show that the analytical methods which most frequently used in the determination of chromium in biomaterials are spectral methods. It complements our rationale for the review of the literature and confirms our practical grounds the necessity of this review.

2. Atomic Spectrum (as) Analytical Methods for the Determination of Trace Chromium in Biomaterials

Traces of chromium in biomaterials are determined by atomic spectrum (AS) analytical methods by FAAS, ETAAS, ICP-OES and ICP-MS. The instrumental detection limits of chrome by FAAS, ETAAS, ICP-OES and ICP-MS are shown in Table 2.

Table 2. Instrumental detection limits (ILOD) chromium by AS – methods.

	FAAS	ETAAS	ICP-OES	ICP-MS	References
ILOD	0.01 ppm	0.1 ppb	0.01 ppm	0.01 ppb	[36]
	50-100 µg/l	0.1 µg/l	1–10 µg/l	0.1µg/l	[1, 37]

In more recent literature is considered that when determining the ILOD chromium by ICP-OES are comparable to those of ET-AAS, and not by FAAS - methods [37]. Emphasizes however, that ILOD in ICP-OES can be aggravated by several times to an order of magnitude in the presence of interfering complex matrices, such as in biological samples [37].

In determining chromium by FAAS require application of the procedures for pre-concentration and / or separation (Table 1 and 2). Thus, by FAAS, after using magnetic nanoparticles as the adsorbent in the solid phase extraction is determined chromium in environmental water and blood serum [14]. By FAAS with pre-concentration, usually by mineralization and / or separation are determined traces of chromium in plasma [24], in hair [27 – 29, 38], in blood [25], liver [25], urine [17, 18], bone [33]. FAAS is used in combination with flow injection analysis, FIA. With FAAS and FIA is determined chromium in urine [18].

In FAAS has a practical limitation, such as the volume of the test sample, which does not allow its use for the determination of chromium in all biological fluids.

ET-AAS is most used method for the determination of trace elements, including chromium in biological samples. Atomization, difference of interfering sources and techniques for their elimination and control in the determination of chromium is given in a general view of a number of specific articles / [1, 15, 16, 39, 40, 41, 42].

For the determination of chromium in ETAAS using preliminary concentration - mineralization and / or extraction mainly in isolated cases deproteinization of blood and serum. It is determined chromium in blood serum samples [1, 5, 12, 13, 43]; blood / [1 - 6, 30, 44]; urine [1, 2, 12, 15 - 17, 19, 34, 42, 45, 46]; milk [1, 21]; liver [26, 44]; hair [2, 30, 31]; bone tissues [4, 5]; nail [1, 15, 16]; erythrocytes [15, 16, 23]; saliva [1, 43]; lung [1]; skin [1]; sweat [1], hip joint tissues [35], marrow [44], kidney [44], and stomach [44]. Have been developed and direct ETAAS methods for the determination of chromium in blood [2]), urine [2, 34, 45], hair [31], blood serum [2].

Upon determination of the chromium and others metals in *ICP-OES* by pretreatment of the samples is used mainly mineralization [7, 22]. Chromium is determined in urine [7] blood [11], nail [7, 22], serum [7], urine [7], hair [7, 11, 22], breast [20], milk [20].

Among the advantages of the method: efficient excitation of the atoms and ions; high stability of discharge; wide linear dynamic range: 5-6 range; ICP-OES - method has some drawbacks: high risk of spectral interferences; direct determination of trace elements in OES, including chromium, amid high concentrations alkali and alkaline earth metals in biological samples is considered to be virtually impossible. Furthermore, the plasma is unstable in the presence of organic substances in biological samples or adding organic solvents [36].

In mass spectroscopy (ICP-MS), the pre-treatment generally requires dilution with nitric acid [32]. The limit of detection of the method in the determination of chromium in unfurnished conditions was 0.01 µg/l, but it does not occur if the total matrix concentration is 1.0 g/l. For comparison, in ETAAS influence matrix concentrations of 20 g /l [47]. So in determinations should be used in several ml samples - 2.5 - 5.0 ml [47] and practical limits of detection of chromium in the biomaterials are in the range 0.1 - 1.0 µg/l [48]. Limitations of the method when working with organic matrix depend on high concentrations of alkali and alkaline earth metals in biological samples.

In ICP-MS chromium is determined in the urine [8], blood serum [8, 10], whole blood [10], and prostate tissue [32].

3. Spectrophotometric Methods for the Determination of Trace Chromium in Biomaterials

Spectrophotometric methods are applied in the

determination of chromium in water analysis [49 - 53]. Another area of application is the analysis of soils, alloy steels, geological samples and industrial effluents [54-57]. A third area of application is the analysis of food and pharmaceutical additives [54, 58, 59]. In comparison to the above methods for the determination of chromium in the analytical sites, few are spectrophotometric method for determination of chromium in human body fluids and tissues. Chromium is determined by spectrophotometry in human blood serum [9, 60-65]. and human urine [9, 60].

The low concentrations of chromium in biological samples (Table 1) require for its determination to apply methods with low limits of detection. Furthermore, AS-methods suitable for the purpose and methods would be suitable chelating agents for chromium, in the micellar environment with spectrophotometric indication. With the help of these methods may be defined concentrations in nanogram range (ng/ml).

We have developed and implemented two methods of chelating agents for chromium in micellar medium and spectrophotometric indication in the determination of chromium in blood serum and urine [66, 67].

In comparison with conventional FAAS and photometric methods for determination of Cr in biological materials, new methods [66, 67] have the following advantages:

- Lower limit of detection, allowing the use of small-volume samples of blood serum and urine - 1.00 ml;

- High selectivity avoids extraction and stripping in the definition used in photometric and flame AAS methods;

- Good reproducibility in determining Cr. In determining the chromium concentration in the range $0.20 - 4.00 \mu mol/l$ in serum samples and urine: relative standard deviation, RSD = 3.24% and RSD = 1.81% on average in the same concentration range for serum and urine, respectively.

- Using the same analytical procedure - reagents, pretreatment of samples, analytical determination in the determination of Cr in blood serum and urine.

- Therefore, both methods can be applied for serial analysis in biomedical laboratories.

The reduced concentration of chromium in blood serum and urine in children with high blood pressure [66, 67] may be considered as a criterion for determining of functional and primary vascular changes in the cardiovascular system.

New methods for the determination of Cr in blood serum and urine are embedded in the Department of "Chemistry and Biochemistry" laboratory Molecular spectrophotometry, elemental and functional bioassay and the Department of Pediatrics of MU-Pleven.

Applications developed methods for solving medicalbiological problems, not casuistry and real cases in medicine, and their implementation in those laboratories showed that methods for determining Cr in blood serum and urine in micellar environment with spectrophotometric indication have a place in modern clinical and toxicological studies.

4. Conclusions

The most ETAAS is used to determine the chromium in

human body fluids and tissues. It is the fastest. In part of the ET-AAS-methods are used direct techniques for determining of chromium in bioassay.

In species with high concentrations of chromium, or those which may be taken larger in volume or mass samples such as hair, nails, urine, is applicable FAAS-method.

ICP-OES and ICP-MS are multi-element methods. In comparison to ET-AAS, the number of studies of chromium in bio samples with them is very measuring less.

AS - methods require expensive and operationally difficult technique with not available most clinical and biomedical laboratories.

Molecular spectral spectrophotometric method is applied in a number of areas to determine the chromium, whereupon the concentrations of chromium in the analytes are comparable to those in human body fluids and tissues. The definitions of chromium, however, in human biosamples in recent years are and in limited numbers limited number analytes. Contemporary applicable methods for the determination of chromium in the biomaterials, by spectrophotometric indication are methods that make use of suitable chelating agents for chromium in the micellar medium. In limit of detection, they are competitive on the AS-methods. And spectrophotometers have all medical and biological laboratories. Furthermore, the methods can be automated and use the available automatic analyzers in clinical laboratories.

We believe that the lag in specifying the chromium concentrations in human bio samples, related to clarifying the role of chromium in health disease can be overcome using both atomic spectral, so spectrophotometric methods. Any determination of chromium in human fluids and tissues, regardless of what method has been made, will contribute as in analytic of the chromium, and in related biomedical research its determination. The available for all medical laboratories spectrophotometric methods are one of the possibilities for participation of more researchers in resolving the analytical, biochemical, physiological, health problems associated with determining of chromium in human body fluids and tissues.

References

- D. L. Tsalev, Atomic Absorption Spectrometry in Occupational and Environmental Health Practice, Vol. III: *Progress in Analytical Methodology, CRC Press, Boca Raton, FL, 1995; ISBN 0-8493-4999-0.*
- [2] Tasneem Gul Kazi, Hassan Imran Afridi, Naveed Kazi, Mohammad Khan Jamali, Mohammad Bilal Arain, Nussarat Jalbani, Ghulam Abbas Kandhro. Copper, Chromium, Manganese, Iron, Nickel, and Zinc, Levels in Biological Samples of Diabetes Mellitus Patients. *Biol Trace Elem Res.* 2008, 122: 1–18.
- [3] Christopher Jantzen, Henrik L Jørgensen, Benn R Duus, Sune L Sporring, and Jes B Lauritzen. Chromium and cobalt ion concentrations in blood and serum following various types of metal-on-metal hip arthroplasties. *Acta Orthop.* 2013, 84(3): 229–236.

- [4] Teresa Lech and Danuta Dudek-Adamska. Optimization and Validation of a Procedure for the Determination of Total Chromiumin Postmortem Material by ETAAS. *Journal of Analytical Toxicology*, 2013, 37: 97–101
- [5] Rim Khlifi, Pablo Olmedo, Fernando Gil, Molka Feki-Tounsi, Amine Chakroun, Ahmed Rebai, Amel Hamza-Chaffai. Blood nickel and chromium levels in association with smoking and occupational exposure among head and neck cancer patients in Tunisia November. *Environmental Monitoring and Assessment.* 2013, 20(11) 8282-8294.
- [6] Li Yong, Kristie C. Armstrong, Royce N. Dansby-Sparks, Nathan A. Carrington, James Q. Chambers, and Zi-Ling Xue. Quantitative Analysis of Trace Chromium in Blood Samples. Combination of the Advanced Oxidation Process with Catalytic Adsorptive Stripping Voltammetry. *Anal Chem.* 2006; 78(21): 7582–7587.
- [7] E. Nasli-Esfahani, F. Faridbod1, B.Larijani1, M.R.Ganjali, P. Norouzi, Trace element analysis of hair, nail, serum and urine of diabetes mellitus patients by inductively coupled plasma atomic emission spectroscopy. *Iranian Journal of Diabetes* and Lipid Disorders. 2011, 10, 1-9.
- [8] Rocha GHO, Steinbach C, Munhoz JR, Madia MAO, Faria JK, Hoeltgebaum D, Barbosar F, Batista BL, Souza VCO, Nerilo SB, Bando E, Mossini SAG, Nishiyama P. Trace metal levels in serum and urine of a population in southern Brazil. *Journal of Trace Elements in Medicine and Biology. 2015*, DOI: http://dx.doi.org/doi:10.1016/j.jtemb.2015.12.005.
- [9] R. Soomro, M. J. Ahmed, and N. Memon. Simple and rapid spectrophotometric determination of trace level chromium using bis (salicylaldehyde) orthophenylenediamine in nonionic micellar media, *Turkish Journal of Chemistry*, 2011, 35 (1): 155–170.
- [10] A. Alimonti, B. Bocca, E. Mannella, F. Petrucci, F. Zennaro, R. Cotichini, C. D'ippolito, A. Agresti, S. Caimi and G. Forte. Assessment of reference values for selected elementsin a healthy urban population. *Ann Ist Super Sanita* 2005; 41(2): 181-187.
- [11] I. O. Olabanji, E. A. Oluyemi, F. O. Fatoye and J. C. Ngila. Elemental composition of blood and hair of mentally–ill patients using ICP-OES. *Techniques. Int. J. Biol. Chem. Sci*, 2011; 5(2): 663-679.
- [12] Nadica Todorovska, Irina Karadjova, Sonja Arpadjan, Trajce Stafilov. On chromium direct ETAAS determination in serum and urine. *Central European Journal of Chemistry*. 2007, 5(1), 230-238.
- [13] Auns Q. Hashim Al-Neami. Measurement of Trace Elements Association with Diabetes Mellitus Based on Atomic Absorption Spectrophotometers. *Advances in Life Science and Technology*, 2014, 24; 21–27.
- [14] Yi-Wei Wu, Jing Zhang, Jun-Feng Liu, Zhen-Li Deng, Mu-Xian Han, Feng Jiang, Dai-Zhi Wang, and Hui-Zhong Yuan. Determination of Chromium Species in Environmental Water and Human Serum Samples by FAAS After Magnetic Solid Phase Extraction. *Atomic spectroscopy*, 2011, 32(1), 43-50.
- [15] R. B. Georgieva, D. L. Tsalev. Chromium levels in erythrocytes, nails and urine as biomarkers of exposure – informational value and relevance. *Bulgarian Journal of Chemistry*, 2014, 3(1): 133-142.
- [16] R.B Georgieva DL Tsalev. Lead, chromium, manganese -

toxicological profile, biological control and analytical determination. *Chemistry and Industry*. 2012, 83, 41-51.

- [17] Joseph M. Makaya, Aly Savadogo, Marius K. Somda, Jean-Baptiste Bour, Nicolas Barro, Alfred S. Traoré. Quality of Human Urine Used as Fertilizer: Case of an Ecological Sanitation Systemin Ouagadougou Peri-Urban Areas-Burkina Faso. Journal of nvironmental Protection, 2014, 5, 467-474.
- [18] Rosa Mari'a Cespo'n, Mari'a Carmen Yebra. Flow injection determination of total chromium in urine of occupationally exposed workers. *Microchim Acta*. 2008, 164: 225–229.
- [19] Benova D, Hadjidekova V, Hristova R, Nikolova T, Boulanova M, Georgieva I, Grigorova M, Popov T, Panev T, Georgieva R, Natarajan AT, Darroudi F and Nilsson R. Cytogenetic effects of hexavalent chromium in Bulgarian chromium platers, *Mutation Research*, 2002, 514: 29-38.
- [20] Nina Bilandžić, M. Sedak, M. Đokić, Đ. Božić. Determination of Macro- and Microelements in Cow, Goat, and Human Milk Using Inductively Coupled Plasma Optical Emission Spectrometry. 2015, 48(9): 677-684.
- [21] Pereira Lara, Paulo Celso, Nicácio Silveira, Josianne, Borges Neto, Waldomiro, Beinner, Mark A., da Silva, José B. B. Use of multivariate optimization to develop a method for direct chromium determination in breast milk by GF-AAS using aqueous calibration. *Journal of Chemical & Pharmaceutical Research.* 2015, 7:(3), 1900-1906.
- [22] A. A. Momen, M. A. A. Khalid1, M. A. A. Elsheikh, D. M. H. Ali, Trace elements in scalp hair and fingernails as biomarkers in clinical studies. *Journal of Health Specialties*. 2015, 3(1): 154-160.
- [23] Zhang J, Li GR, Liu LZ, Zhang N, Wang TC, Yan L, Jia G, Wang X. Chromium content in erythrocytes serving as the exposure biomarker for workers exposed to soluble chromate. *PubMed*. 2006; 40(6): 390-4.
- [24] R.S. Ajibola, O. A. Ogundahunsi, O. O. Soyinka, E. O. Ogunyemi and A. O. Odewabi. Serum Chromium, Molybdenum, Zinc and Magnesium Levels in Diabetes Mellitus Patients in Sagamu, South West Nigeria. *Asian Journal of Medical Sciences*. 2014, 6(2): 15-19.
- [25] Irnius, Danutë Speièienë, Karolina Pajenèkovskytë, Stasys Tautkus, Rolandas Kazlauskas, Aivaras Kareiva. Rapid quantitative determination of metals in blood and liver by FAAS. *Journal of Trace Elements in Medicine and Biology*. 2005, 16(3-4): 29–33.
- [26] Bona MA, Castellano M, Plaza L, Fernandez A. Determination of heavy metals in human liver. *Hum Exp Toxicol.* 1992, 11(5): 311-313.
- [27] Fakayode, S. Owen, D. Pollard and M. Yakubu, "Use of Flame Atomic Absorption Spectroscopy and Multivariate Analysis for the Determination of Trace Elements in Human Scalp," *American Journal of Analytical Chemistry*. 2013, 4(7): 348-359.
- [28] A. Shunmuga Perumal, A. Thangamani. Atomic absorption spectrophotometric determination of heavy metals lead and chromium levels in human hair of people living in katpadi and yelagiri hills of vellore district. *International Journal of Research in Ayurveda & Research Article (IJRAP)*. 2011, 2 (5) 1568-1570.

- [29] Supaporn Pengping and Sukjit Kungwankunakorn. Determination of Some Heavy Metals in Human Hairby Ultrasonic Acid Digestion and Atomic Absorption Spectrophotometry. *Chiang Mai J. Sci.* 2014, 41(1):148-155.
- [30] Fernando Gil, Antonio F. Hernández, Claudia Márquez Pedro Femia c, Pablo Olmedo, Olga López-Guarnido, Antonio Pla. Biomonitorization of cadmium, chromium, manganese, nickel and lead in wholeblood, urine, axillary hair and saliva in an occupationally exposed population. *Science of the Total Environment.* 2011, 409: 1172–1180.
- [31] J. Stupar, F. Dolinsek -Determination of chromium, manganese, lead and cadmium inbiological samples including hair using direct electrothermal atomicabsorption spectrometry. *Spectrochimica Acta Part B 51*. 1996, 665-683.
- [32] Vladimir Zaichick, Sofia Zaichick Use of Neutron Activation Analysis and Inductively Coupled Plasma Mass Spectrometry for the Determination of Trace Elements in Pediatric and Young Adult Prostate. *American Journal of Analytical Chemistry*, 2013, 4, 696-706.
- [33] Brodziak-Dopierała B1, Kwapuliński J2, Sobczyk K3, Wiechuła D1.Chromium content in the human hip joint tissues. *Biomed Environ Sci.* 2015, 28(2):89-96.
- [34] Pietro Apostoli. Graziano Maranelli, Pier Giorgio Duca, Paolo Bavazzano, Angelo Bortoli, Aldo Cruciatti, Giuseppe Elia, Claudio Minoia, Renza Piccinini, Enrico Sabbioni, Gianfranco Sciarra, Claudio Soave. Reference values of urinary chromium in Italy. *Int Arch Occup Environ Health*, 1997, 70: 173-179.
- [35] Brodziak-Dopierała B, Kwapuliński J, Sobczyk K, Wiechuła D. Chromium content in the human hip joint tissues. *Biomed Environ Sci.*, 2015, 28(2):89-96.
- [36] G. Christian and J. O'Reilly. Instrumental Analysis. Publishing University of Sofia, Sofia, 1998.
- [37] D. L. Tsalev, Atomic emission spectrometry with inductivelycoupled plasma in "Fundamentals of chemical analysis" Compiled by prof. DSc Eng. Rahila Borisova, ed. Aquarius, Sofia, 2009, Chapter 4, Section 4.1.2, p. 214-226. ISBN 978-954-9415-43-5.
- [38] Supaporn Pengping and Sukjit Kungwankunakorn. Determination of Some Heavy Metals in Human Hair by Ultrasonic Acid Digestion and Atomic Absorption Spectrophotometry. *Chiang Mai J. Sci.* 2014; 41(1):148-155.
- [39] Dayene C. Carvalho, Nivia M. M. Coelho, Luciana Melo Coelho, Simone S. S. Borges, Thais S. Neri, Vanessa N. Alves. Strategies to increase selectivity of analytical methods for As, Cr and Se speciation in biological samples. *Sample Preparation*. 2014, 2(1):1–12.
- [40] A. G. G. Dionisio, A. M. Dantas de Jesus, R. St'abile Amais, G. Lu'is Donati, Kelber dos Anjos Miranda. Marcelo Braga Bueno Guerra, Joaquim Araujo Nobrega, and Edenir Rodrigues Pereira-Filho. Old and New Flavors of Flame (Furnace) Atomic Absorption Spectrometry. *International Journal of Spectroscopy*. 2011, Article, 30 pages.
- [41] D. L. Tsalev, L. Simeonov, M. Kochubovski, B. Simeonova. Environmental Heavy Metal Pollution and Effects on Child Mental Development, Risk Assessment and Prevention Strategies. *Springer, Dordrecht*, 2011.
- [42] D.C. Carvalho, N. M. M. Coelho, L. Melo Coelho, S. S. S. Borges, Thais S. Neri, V.N. Alves. Strategies to increase

selectivity of analytical methods for As, Cr and Se speciation in biological samples: A review. *Sample Preparation, 2014, 2* (1) 1-12.

- [43] Ağaoğlu, G., Arun, T., İzgü, B., & Yarat, A. Nickel and Chromium Levels in the Saliva and Serum of Patients with Fixed Orthodontic Appliances. *Angle Orthodontist*, 2001, 71 (5):375-379.
- [44] Teresa Lech and Danuta Dudek-Adamska. Optimization and Validation of a Procedure for the Determination of Total Chromium in Postmortem Material by ETAAS. *J Anal Toxicol*, 2013, 37 (2): 97-101.
- [45] Ting-Wan Lin and Shang-Da Huang.Direct and Simultaneous Determination of Copper, Chromium, Aluminum, and Manganese in Urine with a Multielement Graphite Furnace Atomic Absorption Spectrometer. *Anal. Chem.*, 2001, 73 (17), 4319–4325.
- [46] F Barbosa et al. Evaluation of electrodeposited tungsten chemical modifier for direct determination of chromium in urine by ETAAS. *Microchemical Journal*, 2004; 78 (7) 13-19.
- [47] R. Cornelis, B. Heinzow, R. F. M. Herber, M. Jakubowski, J. Molin Christensen, M. Poulsen, E. Sabbioni, D. M. Templeton. Sample collection guidelines for trace elements in blood & urine. *Pure & Appl. Chem*, 1995, 67 (8/9) 1575-1608.
- [48] Vandecasteele, C., and Block, C.B., Modern methods for trace elements determination, John Wiley &Sons1993, Chichester.
- [49] Aggarwal SK, Kinter M., Fitzgerald RL and Herold DA. Masss pectrometry of trace elements in biological samples. *Critical reviews in clinical sciences*, 1994, 31; (1) 35–87.
- [50] M. Llobat-Estellés, A. Maurí-Aucejo, M. López-Catalán. Spectrophotometric determination of chromium with diphenylcarbazide in the presence of vanadium, molybdenum, and iron after separation by solid-phase extraction. *Fresenius' Journal of Analytical Chemistry*, 2001, 371 (3), 358-363.
- [51] P. Nagaraj, N. Aradhana, A.Shivakumar, A. Kumar Shrestha, A. K. Gowda. Spectrophotometric method for the determination of chromium (VI) in water samples. *Environmental Monitoring and Assessment*, 2009, 157, (1-4) 575-582.
- [52] I. Sreevani, P. Raveendra Reddy, V. Krishna Reddy. A Rapid and Simple Spectrophotometric Determination of Traces of Chromium (VI) in Waste Water Samples and in Soil samplesby using 2-Hydroxy, 3-Methoxy Benzaldehyde Thiosemicarbazone (HMBATSC). *IOSR Journal of Applied Physics* (IOSR-JAP), 2013, 3 (1); 40-45.
- [53] Xiaokun Wang, Yingqin Wei, Shasha Wang, Lingxin Chen. Red-to-blue colorimetric detection of chromium via Cr (III)citrat chelating based on Tween 20-stabilized gold nanoparticles. *Colloids and Surfaces A: Physicochem. Eng. Aspects, 2015*, 472; 57–62.
- [54] Abdollahi H. Simultaneous spectrophotometric determination of chromium (VI) and iron (III) with chromogenic mixed reagents by H-point standard addition method and partial least squares regression. *Analytica Chimica Acta*, 2001, 442; (2):327-336.
- [55] Maheswari V. and Balasubramanian N. Spectrophotometric determination of chromium based on ion-pair formation. *Chem. Anal. (Warsaw)*, 1996, 41, 569.

- [56] E V Larionova and K A Bulygina. Simultaneous spectrophotometric determination of chromium (VI) and iron (III) in alloys. *IOP Conf. Ser.: Mater. Sci. Eng.* 81, 2015.
- [57] Martelli P B, Reis B F, Kronka E A M, F H B, Korn M, Zagatto E A G and Araujo A N. Multicommutation in flow analysis. part 2. binary sampling for spectrophotometric determination of nickel, iron and chromium in steel alloys *Analytica Chimica Acta*, 1995, 308 (1), 397-405.
- [58] Luciene S. de Carvalho, Antônio Celso S. Costa, Sérgio L. C. Ferreira, Leonardo S. G. Teixeira. Spectrophotometric determination of chromium in steel with 4-(2- thiazolylazo)resorcinol (TAR) using microwave radiation. J. Braz. Chem. Soc., São Paulo, 2004, 15(1):153-157.
- [59] C. Tayone. Spectrophotometric Determination of Chromium (VI) in Canned Fruit Juices J. International Journal of Sciences: Basic and Applied Research (IJSBAR), 2015, 19(1): 426-432.
- [60] De, AK (2003). Environmental Chemistry, 5th ed.; New Age International, New Delhi.
- [61] Quanti Chrom TM, Chromium Assay Kit (DCRM-250) Quantitative Colorimetric Determination of Chromium (2012, in online). BioAssay Systems 3191, Corporate Place Hayward, CA 94545 USA.

- [62] Fabiyi, F.A.S., Donnio, A.Z. Use of variamine blue as a chromogenic reagent for rapid spectrophotometric determination of nano amount of chromium. *Synthesis and Reactivity in Inorganic, Metal-Organic and Nano-Metal Chemistry*, 2007, 37(10):809-812.
- [63] D. G. Barceloux. Chromium. J Toxicol Clin Toxicol, 1999, 37(2): 173-94.
- [64] Chromium Assay Kit (MAK130) Technical Bulletin Sigma -Aldrich. Colorimetric Test of Chromium (2013, in online). http://www.sigmaaldrich.com/content/dam/sigmaaldrich/docs/Sigma/Bulletin/2/mak130bul.pdf
- [65] Abnova[™] Chromium Assay Kit (Colorimetric). Manufacturer: ABNOVA CORPORATION KA3771. Cat. No. 89-101-188. Sample Type Beverage, Food, Plasma, Serum, Soil, Water. https://www.fishersci.com/shop/products/abnovachromium-assay-kit-colorimetric-1-kit/89101188
- [66] Maria G. Angelova, Atanaska N. Bozhinova, Nadia Kolarova-Ianeva. Determination of serum chromium and application at children with primary arterial hypertension. *American Journalof Chemistry and Applications*, 2015, 2 (1): 1-4.
- [67] Atanaska N. Bozhinova, Maria G. Angelova. Trace Chromium in Human Urine Samples with Photometric Detection Method. *American Journal of Chemistry and Applications*, 2015; 2(3): 61-65.